

## TiO<sub>2</sub>/Chitosan bioplastic as Antibacterial of *Stephylococcus aureus* for Food Preservation

Indar Kustiningsih<sup>1\*</sup>, Dhena Ria Barleany<sup>1</sup>, Devi Abriyani<sup>1</sup>, Asep Ridwan, Muhammad Syairazy, Mochamad Adha Firdaus

<sup>1</sup>Chemical Engineering Department Sultan Ageng Tirtayasa University, Cilegon, Indonesia

<sup>2</sup>Chemical Engineering Department King Fahd University of Petroleum and Minerals, Dhanran, Saudi Arabia

\*Corresponding Author Email: [indar.kustiningsih@untirta.ac.id](mailto:indar.kustiningsih@untirta.ac.id)

### ARTICLE HISTORY

Received April 7<sup>th</sup>, 2021  
Received in revised form May 19<sup>th</sup>, 2021  
Accepted June 22<sup>nd</sup>, 2021  
Available online September 1<sup>st</sup>, 2021

### ABSTRACT

Nowadays, bioplastic development become hot trends to assess environmental issues. Many materials have been purposed to be the best resources for bioplastic manufacturing. Chitosan is one of the most abundant resources in which could derivates from biomaterial waste called chitin. TiO<sub>2</sub> nanoparticles incorporation within biomaterial presumably not only enhance its mechanical properties but also improve biocompatibility of medical characteristic such as bacterial annihilation. From this study, it was shown that small amount of TiO<sub>2</sub> nanoparticles within chitosan bioplastic prove improvement of both characteristic. Nevertheless, it was also slightly increasing material durability to degrade.

**Keywords:** TiO<sub>2</sub>, bioplastic, Antibacterial, *Stephylococcus aureus*

### 1. INTRODUCTION

Recent development of biodegradable plastic manufacturing for food preservation to also solve environmental issues aimed from second derivatives of natural ingredients like chitosan (Diaz, V. et al., 2010; Fathanah, dkk., 2018; Ginting, dkk., 2018; Naito et al., 2016; Nishiyama et al., 1996; Ogawa, et el., 2019; Rohmawati, dkk., 2018, Kustiningsih, I., et. al., 2019). Chitosan is one of the biggest natural biopolymer sources after cellulose (Mallakpour & Madani, 2015) that have excellent biodegradable features, nontoxic, and biocompatible (Haldorai & Shim, 2014; Mazin C., 2015; Nikkhoo et al., 2018). Chitosan also have anti-microbial properties and easy to produce with low price (Kashif & Park, 2019; Logpriya et al., 2018; Panariello, Coltelli, Buchignani, & Lazzeri, 2019). Compare to conventional plastic, chitosan has a back draw predominantly in its mechanical properties such as tensile strength and elongation (Mallakpour & Madani, 2015)..

Manufacturing nanocomposite chitosan with nanofiller such as silica, clay, and TiO<sub>2</sub> particles predominantly tended to improve not only mechanical

properties but also enhance biomedical features within nanocomposite material specifically for food preservation (de Azeredo, 2009; Kanmani & Rhim, 2014a; Rhim, 2011; Rhim, Park, & Ha, 2013). Nowadays, nanofiller classified into several specific type based on its shape, including: nanoparticles (Kanmani & Rhim, 2014, Kustiningsih, I., et al., 2019. ), nanofibril (Rafeian, Shahedi, Keramat, & Simonsen, 2014) and nanotubes ((Diaz-Visurraga et al., 2010). Nanoparticle TiO<sub>2</sub> incorporation within chitosan material proven to alter its antimicrobial apart from other nanofiller by growth-control and invasion of bacteria that leads to elimination of pathogenic microorganism (de Azeredo, 2009).

TiO<sub>2</sub> has been widely developed for its medical features such as virus, fungi, algae and cancer cell elimination (Chawengkijwanich & Hayata, 2008; Tsuang et al., 2008). Furthermore, TiO<sub>2</sub> proven could annihilates several bacteria (*Pseudomonas aeruginosa* and *Enterococcus faecalis* bacteria (Jeffery, Pepler, Lima, & McDonald, 2010); *Lactobacillus helveticus* (Liu & Yang, 2003)). Furthermore, there are few reports on the observation of pure nanoparticles TiO<sub>2</sub> for specified *Staphylococcus aureus* elimination within chitosan bioplastic. The experiment of manufacturing

TiO<sub>2</sub>/chitosan bioplastic and its application specifically for food preservation will lead to the opportunity of advancement bio composite material.

## 2. METHODS

One sample of 1-gram chitosan mixed with 1% (v/v) acetic acid glacial, stirred for 3 hours and homogenized for 30 minutes at room temperature. Followed by TiO<sub>2</sub> incorporation for each variation (TiO<sub>2</sub>: 0 g, 0.1 g, 0.2 g, 0.5 g and 1 g), then stirred for 4 hours and homogenized for 1 hour. A solution molded into glass plate and dried at 80°C afterwards.

The characterization of bioplastic was done using Hitachi SU-3500 Scanning Electron Microscope to assess surface morphology, shape smoothness and TiO<sub>2</sub> particle dispersion. Prestige Shimadzu Fourier Transform Infra-Red 8201 was used for chemical functional groups analysis and Universal Testing Machine to analyze mechanical properties such as elongation and tensile strength.

Sterilized nutrient agar medium has been mixed with sample bioplastic. Followed by blending with rejuvenated *Staphylococcus aureus* then dissolved with 103 and 104 solution. Thereafter, plating the mixture upon petri dish for 24 hours incubation for different environment: dark and under UV radiation. Bacteria colony counted afterwards by colony forming units (CFU).

## 3. RESULT and DISCUSSION

### 3.1. Spectrophotometer FTIR

Fourier Transform Infrared Spectroscopy have observed functional groups analysis involving: O-H bond, N-H bond, C=O bond, C-O bond, and C-N bond for pure chitosan bioplastic sample whereas R-NH-R bond and Cs-TiO<sub>2</sub> were detected for TiO<sub>2</sub>/chitosan bioplastic (Figure 1).

The graph shows similarity between pure chitosan bioplastic sample and TiO<sub>2</sub> incorporated chitosan bioplastic. Characteristics transformation of transmittance peak from FTIR analysis occurred as a result from pure chitosan bioplastic mixed with TiO<sub>2</sub> nanoparticles. The mixture has not only physical interaction but also chemical bond (Bourtoom & Chinnan, 2008). Compare to pure chitosan bioplastic, there were shifting of hydroxyl bonds, amino bonds, and amide bonds of TiO<sub>2</sub> incorporated chitosan bioplastic. The changes caused by interaction between chitosan molecules and TiO<sub>2</sub> nanoparticles (Haldorai & Shim, 2014) which is shown by chitosan – TiO<sub>2</sub> bonds on 450 – 950 wavelength range (Amir, Julkapli, & Hamid, 2016).

The spectrum shows that pure chitosan bioplastic has an O-H bonds and N-H bonds at the peak of 3419 cm<sup>-1</sup> whereas functional groups of C-H exhibited at 2924 cm<sup>-1</sup>, C=O at 1616 cm<sup>-1</sup>, C-O at 1257 cm<sup>-1</sup> and C-N at 1039 cm<sup>-1</sup> respectively. On the other hand, peak spectrum of TiO<sub>2</sub> incorporated chitosan bioplastic is a combination of each pure peak compound's chitosan and TiO<sub>2</sub>. It could be said with certainty that there was a cross linkage between chitosan and TiO<sub>2</sub> functional groups as for example the overlapped O-H bonds with N-H bonds at 3439 cm<sup>-1</sup>, R-NH-R at 1600 cm<sup>-1</sup>, O-H deformation at 1442 cm<sup>-1</sup>, Chitosan/TiO<sub>2</sub> deformation at 947 cm<sup>-1</sup>.

### 3.2. Surface layer analysis

The results of Scanning Electron Microscope (SEM) describes the difference upon surface layer of pure chitosan bioplastic and TiO<sub>2</sub> incorporated chitosan bioplastic for each composition (Figure 2). The morphology of pure chitosan bioplastic displays such a homogenous, sleek and clean without any speckles. However, TiO<sub>2</sub> nanoparticles enhancement aimed chitosan bioplastic have a rough structure surface with many grains layered upon.

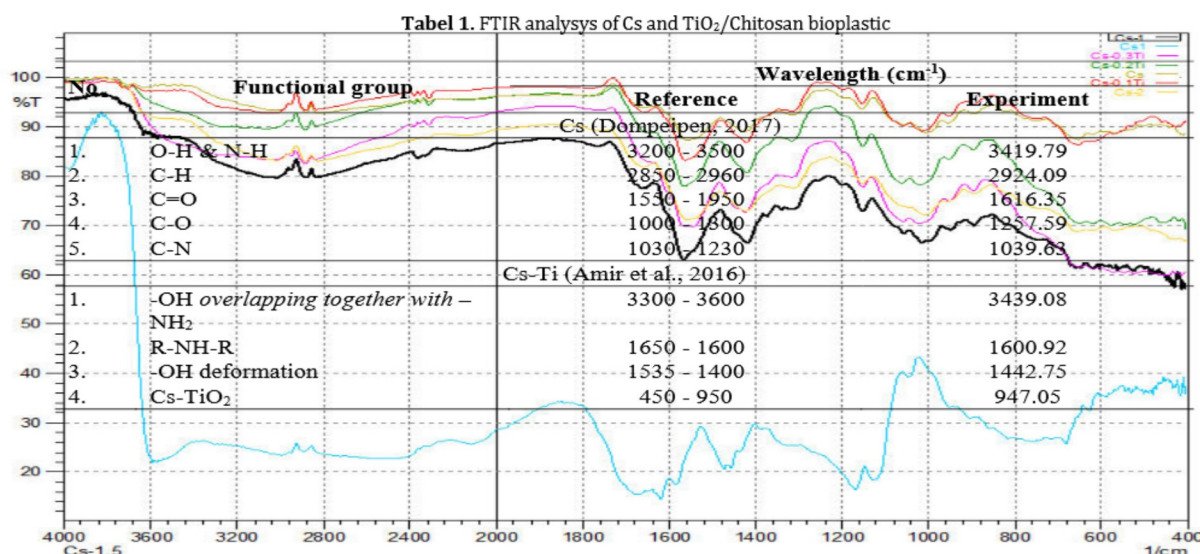


Figure 1. FTIR spectrum of Cs dan TiO<sub>2</sub>/chitosan bioplastic

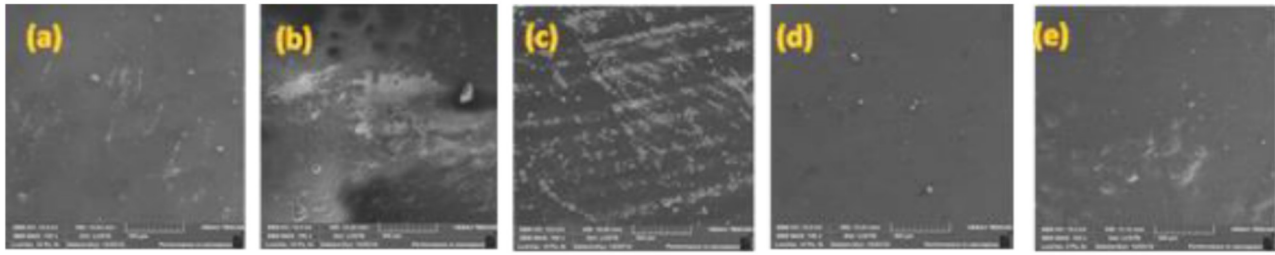


Fig. 2. . Scanning Electron Microscope (SEM) image of chitosan bioplastic and TiO<sub>2</sub> incorporate chitosan bioplastic (a) Cs (b) Cs-OH (c) Cs-0.1 Ti (d) Cs-0.2 Ti (e) Cs-0.3 Ti.

These speckles reveal TiO<sub>2</sub> nanoparticles dispersion within chitosan bioplastic matrix (Figure 2b to Figure 2e). TiO<sub>2</sub> nanoparticles started to exhibit regular pattern when the composition of TiO<sub>2</sub> at 1 : 0.1 within chitosan bioplastic (Figure 2c). At ratio of 1 : 0.2 and 1 : 0.3 (Cs : TiO<sub>2</sub>), the distribution of TiO<sub>2</sub> nanoparticles within chitosan bioplastic manifests such a fine cluster compare to the chitosan bioplastic surface (Figure 2d and Figure 2e). A proper distribution of TiO<sub>2</sub> nanoparticles within matrix polymer is one of key indicator of a good TiO<sub>2</sub> incorporated chitosan bioplastic (Diaz-Visurraga et al., 2010).

### 3.3. Mechanical Properties analysis (Tensile Strength and Elongation)

Tensile strength and elongation tests are involved on mechanical property analysis of chitosan bioplastic and TiO<sub>2</sub> incorporated chitosan bioplastic (Table 2, Figure 3). TiO<sub>2</sub> enhancement on chitosan bioplastic reduce the both of mechanical features. The amount of TiO<sub>2</sub> nanoparticles composition within chitosan bioplastic change the matrix from homogenous into heterogeneous as appears upon surface layer. The phenomena occurred caused by the irregularity of TiO<sub>2</sub> nanoparticles distribution within chitosan bioplastic matrix as shown

in Figure 2b to Figure 2e. These finding also align from others that discovered the effect of TiO<sub>2</sub> nanoparticles on mechanical properties of chitosan (Mazin C., 2015).

Enhancement of TiO<sub>2</sub> nanoparticles within chitosan bioplastic gave a positive results of mechanical properties improvement. For a sample with 0.1 g TiO<sub>2</sub> composition, tensile strength exhibited is 16.93 MPa (decreasing 10.55 MPa) with observed elongation is 8.67 % (increasing 6.02%) from control sample which made from pure chitosan bioplastic. Furthermore, for 0.2 g TiO<sub>2</sub> composition, observed tensile strength is 29.46 (increasing 1.98 MPa) with elongation exhibited was 8.67 % (same with 0.1 g TiO<sub>2</sub>).

Afterwards, increasing TiO<sub>2</sub> nanoparticles into chitosan bioplastic mixture seems to be declining its mechanical properties. It was shown that for TiO<sub>2</sub> composition at 0.3 g, 1 g, 2 g and 3 g, all of them have smaller mechanical properties rather than control sample of pure chitosan. These phenomena happened due to influence of TiO<sub>2</sub> nanoparticles that made a change in matrix structure of chitosan. Pure chitosan bioplastic has a homogeneous matrix structure compare to TiO<sub>2</sub> incorporated chitosan bioplastic (Figure 2a to Figure 2e).

Table 2. Influence of TiO<sub>2</sub> nanoparticles to mechanical properties chitosan bioplastic

No	Mixture	Tensile Strength (MPa)	Elongasi (%)
1	Cs	27.48	2.65
2	Cs-1	2.07	3.33
3	Cs-1.5	4.79	2.33
4	Cs-2	11.93	5.33
5	Cs-0.1 Ti	16.93	8.67
6	Cs-0.2 Ti	29.46	8.67
7	Cs-0.3 Ti	15.39	2.33

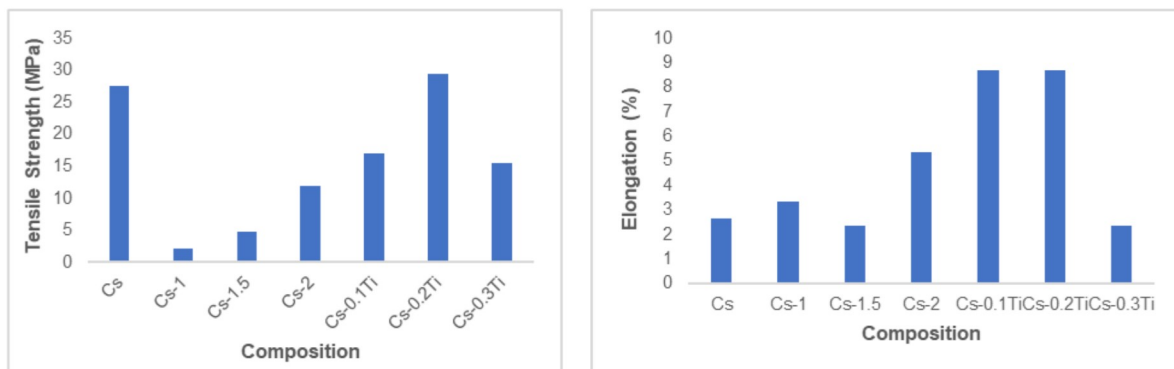


Fig. 3 Mechanical properties analysis (a) Tensile strength; (b) Elongation

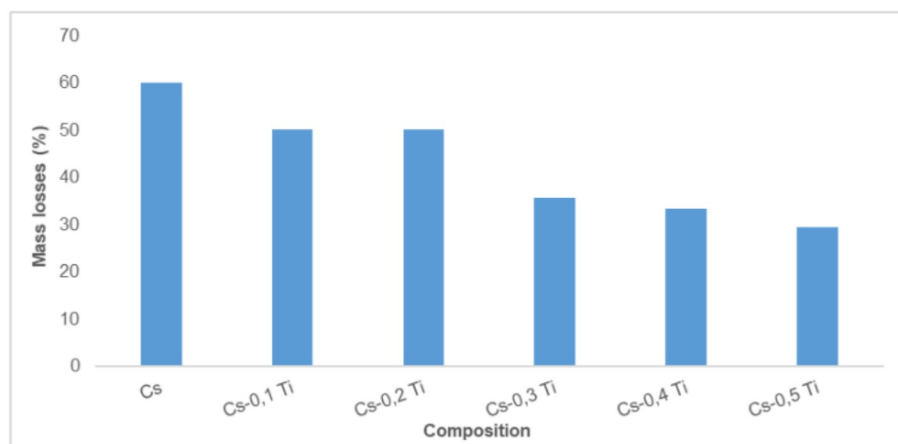
### 3.4. Biodegradation and Antibacterial activity analysis

Biodegradation analysis results after 2 months burial is shown in Table 3. TiO<sub>2</sub> enhancement did not inhibit degradation process of chitosan bioplastic. Mass reducing of bioplastic sample still exhibited due to natural ingredient of bioplastic material (Figure 4). Mass reducing showed up because natural ingredient prone to

easier to be digested by several microorganism. Thus, the more TiO<sub>2</sub> nanoparticles enhanced within chitosan bioplastic apt to prevent the process that leads to the longer time required for degradation. This antimicrobial activity from TiO<sub>2</sub> nanoparticles is become one of the features that could effectively prolong chitosan bioplastic endurance from aging.

**Table 3.** Influence of TiO<sub>2</sub> nanoparticles to mass losses of chitosan bioplastic and TiO<sub>2</sub>/chitosan

Sample	Initial weight (g)	Final weight (g)	Mass losses (%)
Cs	0.05	0.02	60.00
Cs-0,1 Ti	0.10	0.05	50.00
Cs-0,2 Ti	0.10	0.05	50.00
Cs-0,3 Ti	0.14	0.09	35.071
Cs-0,4 Ti	0.15	0.10	33.33
Cs-0,5 Ti	0.17	0.12	29.41



**Figure 4** Biodegradation analysis of chitosan bioplastic and TiO<sub>2</sub>/chitosan bioplastic

Photocatalytic TiO<sub>2</sub> by UV irradiation has been done to verify antibacterial bioplastic characteristic in order to annihilate gram-negative bacteria and gram-positive bacteria including: endospore, fungi, algae, protozoa and also virus (Paspaltsis et al., 2006). Staphylococcus aureus has been used as a representation of gram-positive bacteria. The results of antibacterial activity presented in Table 4.

As by nature, chitosan bioplastic has an antibacterial activity features but not to eliminate the numbers of Staphylococcus aureus. The antibacterial properties come from amino functional groups that has positive charge which could interact with membrane cell of microorganism which has negative charge. This interaction would cause a destruction of protein and other intracellular constituent from microbes. Chitosan has been tested effectively eradicate gram-negative bacteria rather than gram-positive bacteria (Diaz-Visurraga et al., 2010; Zheng et al., 2000)

It has been proven by these experiment and the results that shown in Table 4, TiO<sub>2</sub> nanoparticles enhancement could significantly reduce the amount of

s.aureus bacteria that has been incubated for 24 hours. Interestingly, elimination of bacteria s.aureus could be done either by irradiation or without irradiation by UV lights. These kinds of characteristics also observed in other experiments (Diaz-Visurraga et al., 2010; Tsuang et al., 2008). Photocatalytic activity of TiO<sub>2</sub> nanoparticles can be initiated by UV irradiation that will release positive ion charge within chitosan bioplastic. These positive ion charge then will interact with lipid membrane of s.aureus bacteria in which have negative ion.

Under UV irradiation, TiO<sub>2</sub> nanoparticles will generate electron (e<sup>-</sup>) on conduction band and exhibit the hole (h<sup>+</sup>) on valence band. The hole will interact with water molecule to produce reactive radical hydroxyl (·OH) and ·O<sub>2</sub> which lead to degradation of any organic substance like membrane barrier of cell bacteria (Gumiero et al., 2013). Without UV irradiation, the mechanism of s.aureus bacteria elimination within TiO<sub>2</sub> nanoparticles incorporated mixture remains unknown. It was presumably done by similar process in which Ag

elements become an antibacterial agent. Both of Ag and TiO<sub>2</sub> have a positive charge that can interact with negative charge which comes from membrane barrier of cell bacteria (Amrulia, 2012). After 24 hours incubation, the number of control variable bacteria tended to be decreased (Figure 6) for both methods (photocatalytic

and non-photocatalytic). On pure chitosan bioplastic, bacteria only occurred in several colony. In addition, for every sample tested using TiO<sub>2</sub>/chitosan bioplastic there is none of Staphylococcus aureus bacteria prone to be lived.

Table 4 Antibacterial activity analysis

Sample	UV irradiation		Without radiation	
	CFU (mL <sup>-1</sup> )	Survival Ratio (%)	CFU (mL <sup>-1</sup> )	Survival Ratio (%)
Control	2.9 x 10 <sup>5</sup>	100.0	2.9 x 10 <sup>5</sup>	100.0
Cs	2.0 x 10 <sup>4</sup>	6.9	2.0 x 10 <sup>4</sup>	3.5
TiO <sub>2</sub>	0.0	0.0	0.0	0.0
Cs-0,1 Ti	0.0	0.0	0.0	0.0
Cs-0,2 Ti	0.0	0.0	0.0	0.0
Cs-0,3 Ti	0.0	0.0	0.0	0.0
Cs-0,4 Ti	0.0	0.0	0.0	0.0
Cs-0,5 Ti	0.0	0.0	0.0	0.0

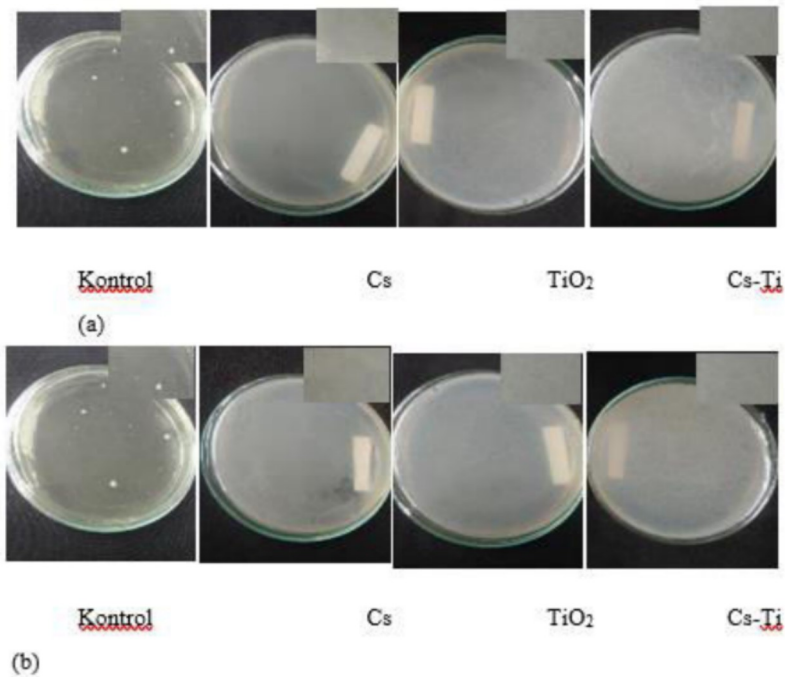


Figure 5. Antibacteria analysis (a) non photocatalytic (b) photocatalytic under UV

#### 4. CONCLUSIONS

TiO<sub>2</sub> incorporation within chitosan bioplastic is viable to be manufactured. For specified application of food preservation, TiO<sub>2</sub> nanoparticles enhancement exhibits improvement of not only mechanical properties features such as tensile strength and elongation but also able to annihilates the number of Staphylococcus aureus bacteria to zero either by using photocatalytic or non-photocatalytic method. Nano filling TiO<sub>2</sub> within chitosan bioplastic also made material slightly more durable to degradation compare to conventional plastic.

## 5. ACKNOWLEDGMENTS

The authors would like to thank to the Islamic Development Bank for financial assistance under IDB-UNTIRTA Research Grant project No. 593/UN43.9/PL/2019.

## 6. REFERENCES

- Amir, M. N. I., Julkapli, N. M., & Hamid, S. B. A. (2016). Incorporation of chitosan and glass substrate for improvement in adsorption, separation, and stability of TiO<sub>2</sub> photodegradation. *International Journal of Environmental Science and Technology*, 13(3), 865-874. doi:10.1007/s13762-015-0914-y
- Amrulia, W. (2012). Uji Aktivitas Antibakteri Kitosan-TiO<sub>2</sub> Pada Tekstil Terhadap *Escherichia coli*. [Antibacteria activity test of chitosan - TiO<sub>2</sub> on textile material for *Escherichia coli*].
- Bourtoom, T., & Chinnan, M. S. (2008). Preparation and properties of rice starch-chitosan blend biodegradable film. *Lwt-Food Science and Technology*, 41(9), 1633-1641. doi:10.1016/j.lwt.2007.10.014
- Cai, R. X., Kubota, Y., Shuin, T., Sakai, H., Hashimoto, K., & Fujishima, A. (1992). Induction of Cytotoxicity by Photoexcited TiO<sub>2</sub> Particles. *Cancer Research*, 52(8), 2346-2348. Retrieved from <Go to ISI>://WOS:A1992HN84200036
- Chawengkijwanich, C., & Hayata, Y. (2008). Development of TiO<sub>2</sub> powder-coated food packaging film and its ability to inactivate *Escherichia coli* in vitro and in actual tests. *International Journal of Food Microbiology*, 123(3), 288-292. doi:10.1016/j.jfoodmicro.2007.12.017
- de Azeredo, H. M. C. (2009). Nanocomposites for food packaging applications. *Food Research International*, 42(9), 1240-1253. doi:10.1016/j.foodres.2009.03.019
- Diaz-Visurraga, J., Melendrez, M. F., Garcia, A., Paulraj, M., & Cardenas, G. (2010). Semitransparent Chitosan-TiO<sub>2</sub> Nanotubes Composite Film for Food Package Applications. *Journal of Applied Polymer Science*, 116(6), 3503-3515. doi:10.1002/app.31881
- Dompeipen, E. J. (2017). Isolasi dan identifikasi kitim dan kitosan dari kulit udang windu (*Penaeus monodon*) dengan spektroskopi inframerah. *Majalah BIAM*, (2017), 12.
- Fathanah, U., Lubis, M. R., Nasution, F., & Masyawi, M. S. (2018). Characterization of bioplastic based from cassava crisp home industrial waste incorporated with chitosan and liquid smoke. 3rd International Conference on Chemical Engineering Sciences and Applications 2017 (3rd Ichesa 2017), 334. doi:Unsp 012073 10.1088/1757-899x/334/1/012073
- Ginting, M. H. S., Lubis, M., Sidabutar, T., & Sirait, T. P. (2018). The effect of increasing chitosan on the characteristics of bioplastic from starch talas (*Colocasia esculenta*) using plasticizer sorbitol. *Friendly City 4 from Research to Implementation for Better Sustainability*, 126. doi:Unsp 012147 10.1088/1755-1315/126/1/012147
- Gumiero, M., Peressini, D., Pizzariello, A., Sensidoni, A., Iacumin, L., Comi, G., & Toniolo, R. (2013). Effect of TiO<sub>2</sub> photocatalytic activity in a HDPE-based food packaging on the structural and microbiological stability of a short-ripened cheese. *Food Chemistry*, 138(2-3), 1633-1640. doi:10.1016/j.foodchem.2012.10.139
- Haldorai, Y., & Shim, J. J. (2014). Novel Chitosan-TiO<sub>2</sub> Nanohybrid: Preparation, Characterization, Antibacterial, and Photocatalytic Properties. *Polymer Composites*, 35(2), 327-333. doi:10.1002/pc.22665
- Jeffery, B., Pepler, M., Lima, R. S., & McDonald, A. (2010). Bactericidal Effects of HVOF-Sprayed Nanostructured TiO<sub>2</sub> on *Pseudomonas aeruginosa*. *Journal of Thermal Spray Technology*, 19(1-2), 344-349. doi:10.1007/s11666-009-9369-3
- Kanmani, P., & Rhim, J. W. (2014a). Physicochemical properties of gelatin/silver nanoparticle antimicrobial composite films. *Food Chemistry*, 148, 162-169. doi:10.1016/j.foodchem.2013.10.047
- Kanmani, P., & Rhim, J. W. (2014b). Properties and characterization of bionanocomposite films prepared with various biopolymers and ZnO nanoparticles. *Carbohydrate Polymers*, 106, 190-199. doi:10.1016/j.carbpol.2014.02.007
- Kashif, S. A., & Park, J. K. (2019). Enzymatically Hydrolyzed Water-Soluble Chitosan as a Potent Anti-Microbial Agent. *Macromolecular Research*, 27(6), 551-557. doi:10.1007/s13233-019-7095-3
- Kustiningsih I, Ridwan A, Abriyani D, Syairazy M, Kurniawan T, Barleany D. R. Development of Chitosan-TiO<sub>2</sub> Nanocomposite for Packaging Film and its Ability to Inactive *Staphylococcus Aureus*. *Orient J Chem* 2019;35(3). Available from: <https://bit.ly/2MUXwF1>
- Liu, H. L., & Yang, T. C. K. (2003). Photocatalytic inactivation of *Escherichia coli* and *Lactobacillus helveticus* by ZnO and TiO<sub>2</sub> activated with ultraviolet light. *Process Biochemistry*, 39(4), 475-481. doi:10.1016/S0032-9592(03)00084-0
- Logpriya, S., Bhuvaneshwari, V., Vaidehi, D., SenthilKumar, R. P., Malar, R. S. N., Sheetal, B. P., . . . Kalaiselvi, M. (2018). Preparation and characterization of ascorbic acid-mediated chitosan-copper oxide nanocomposite for anti-microbial, sporicidal and biofilm-inhibitory activity. *Journal of Nanostructure in Chemistry*, 8(3), 301-309. doi:10.1007/s40097-018-0273-6
- Mallakpour, S., & Madani, M. (2015). Effect of Functionalized TiO<sub>2</sub> on Mechanical, Thermal and Swelling Properties of Chitosan-Based Nanocomposite Films. *Polymer-Plastics Technology and Engineering*, 54(10), 1035-1042. doi:10.1080/03602559.2014.974194
- Mazin C., T., A., Anandapadmanabhan, Ashfaq, Mujeeb, A., dan Lobo, A. G. (2015). Study on the Effect of Nano TiO<sub>2</sub> on Mechanical Properties of Chitosan. *IOSR Journal of Mechanical and Civil Engineering*, 12(3), 7. doi:10.9790/1684-12314854
- Naito, P. K., Ogawa, Y., Sawada, D., Nishiyama, Y., Iwata, T., & Wada, M. (2016). X-ray Crystal Structure of Anhydrous Chitosan at Atomic Resolution. *Biopolymers*, 105(7), 361-368. doi:10.1002/bip.22818
- Nikkhoo, M., Amini, M., Farnia, S. M. F., Mandavinia, G. R., Gautam, S., & Chae, K. H. (2018). Preparation and Characterization of Magnetic Chitosan/Cu-Mg-Al Layered Double Hydroxide Nanocomposite for the One-Pot Three-Component (A<sub>3</sub>) Coupling of Aldehydes, Amines and Alkynes. *Journal of Inorganic and Organometallic Polymers and Materials*, 28(5), 2028-2035. doi:10.1007/s10904-018-0861-4
- Nishiyama, M., Hosokawa, J., Yoshihara, K., Kubo, T., Kabeya, H., Endo, T., & Kitagawa, R. (1996). Biodegradable plastics derived from cellulose fiber and chitosan. *Hydrophilic Polymers*, 248, 113-123. Retrieved from <Go to ISI>://WOS:A1996BE65L00007
- Ogawa, Y., Naito, P. K., & Nishiyama, Y. (2019). Hydrogen-bonding network in anhydrous chitosan from neutron crystallography and periodic density functional theory calculations. *Carbohydrate Polymers*, 207, 211-217. doi:10.1016/j.carbpol.2018.11.042
- Panariello, L., Coltelli, M. B., Buchignani, M., & Lazzeri, A. (2019). Chitosan and nano-structured chitin for biobased anti-microbial treatments onto cellulose based materials. *European Polymer Journal*, 113, 328-339. doi:10.1016/j.eurpolymj.2019.02.004
- Paspaltsis, I., Kotta, K., Lagoudaki, R., Grigoriadis, N., Poullos, I., & Sklaviadis, T. (2006). Titanium dioxide photocatalytic inactivation of prions. *Journal of General Virology*, 87, 3125-3130. doi:10.1099/vir.0.81746-0
- Rafieian, F., Shahedi, M., Keramat, J., & Simonsen, J. (2014). Mechanical, thermal and barrier properties of nano-biocomposite based on gluten and carboxylated cellulose nanocrystals. *Industrial Crops and Products*, 53, 282-288. doi:10.1016/j.indcrop.2013.12.016
- Rhim, J. W. (2011). Effect of clay contents on mechanical and water vapor barrier properties of agar-based nanocomposite films. *Carbohydrate Polymers*, 86(2), 691-699. doi:10.1016/j.carbpol.2011.05.010
- Rhim, J. W., Park, H. M., & Ha, C. S. (2013). Bio-nanocomposites for food packaging applications. *Progress in Polymer Science*, 38(10-11), 1629-1652. doi:10.1016/j.progpolymsci.2013.05.008
- Rohmawati, B., Sya'idah, F. A. N., Rhismayanti, Alighiri, D., & Eden, W. T. (2018). Synthesis of Bioplastic-based Renewable Cellulose Acetate from Teak Wood (*Tectona grandis*) Biowaste Using Glycerol-Chitosan Plasticizer. *Oriental Journal of Chemistry*, 34(4), 1810-1816. doi:10.13005/ojc/3404014
- Tsuang, Y. H., Sun, J. S., Huang, Y. C., Lu, C. H., Chang, W. H. S., & Wang, C. C. (2008). Studies of photokilling of bacteria using titanium dioxide nanoparticles. *Artificial Organs*, 32(2), 167-174. doi:10.1111/j.1525-1594.2007.00530.x
- Zheng, H., Maness, P. C., Blake, D. M., Wolfrum, E. J., Smolinski, S. L., & Jacoby, W. A. (2000). Bactericidal mode of titanium dioxide photocatalysis. *Journal of Photochemistry and Photobiology a-*

Chemistry, 130(2-3), 163-170. doi:Doi 10.1016/S1010-6030(99)00205-1